Blood Plasma Proteins, Coagulation and Fibrinolysis

The blood is the body's main transport system. Although the transport and delivery of oxygen to the cells of the tissues is carried out by specialized cells, other vital components such as nutrients, metabolites, electrolytes, and hormones, are all carried in the noncellular fraction of the blood, the plasma. Some components, such as glucose, are dissolved in the plasma; others, for example, lipids and steroid hormones, are bound to carrier proteins for transport. The osmotic pressure of the plasma proteins regulates the distribution of water between the blood and the tissues. Plasma proteins in conjunction with platelets maintain the integrity of the circulatory system through the process of clotting.

Blood normally circulates through endothelium-lined blood vessels. When a blood vessel is severed, a blood clot (called a thrombus, which is formed by the process of thrombosis) forms as part of hemostasis, the physiologic response that stops bleeding. Blood clots also form to repair damage to the endothelial lining, in which components of the subendothelial layer become accessible to plasma proteins.

In hemostasis and thrombosis, a primary hemostatic plug, consisting of aggregated platelets and a fibrin clot, forms at the site of injury. Platelets are attached to the subendothelial layer of the vessel principally through the protein von Willebrand factor, and are activated to bind fibrinogen. Fibrinogen binds the platelets to each other to form a platelet aggregate. The platelet aggregate is rapidly covered with layers of fibrin, which are formed from fibrinogen by the proteolytic enzyme thrombin.

Injury to the endothelium and exposure of tissue factor on the subendothelial layer to plasma proteins also activate the blood coagulation cascade, which ultimately activates thrombin and Factor XIII. Factor XIII cross-links strands of polymerized fibrin monomers to form a stable clot (the secondary hemostatic plug). The blood coagulation cascade consists of a series of enzymes (such as Factor X), which are inactive until proteolytically cleaved by the preceding enzyme in the cascade. Other proteins (Factor V and Factor VIII) serve as binding proteins, which assemble factor complexes at the site of injury. Ca^{2+} and γ-carboxyglutamate residues in the proteins (formed by a vitamin K–dependent process in the liver) attach the factor complexes to phospholipids exposed on platelet membranes. Consequently, thrombus formation is rapidly accelerated and localized to the site of injury.

Regulatory mechanisms within the blood coagulation cascade and antifibrinolytic mechanisms prevent random coagulation within blood vessels that might obstruct blood flow. Impairments in these mechanisms lead to thrombosis.
The hydrostatic pressure in an arteriole is the force that “pushes” fluid out of the capillary and into the interstitial spaces. The plasma protein osmotic pressure, plus the tissue pressure, is the force that “pulls” water from interstitial spaces into the venular side of the capillary. Thus, if the hydrostatic pressure is greater than the osmotic pressure, fluid will leave the circulation; if it is less, fluid will enter the circulation.

In cases of severe protein malnutrition (kwashiorkor), the concentration of the plasma proteins decreases, as a result of which the osmotic pressure of the blood decreases. As a result, fluid is not drawn back to the blood and instead accumulates in the interstitial space (edema). The distended bellies of famine victims are the result of fluid accumulation in the extravascular tissues because of the severely decreased concentration of plasma proteins, particularly albumin. Albumin synthesis decreases fairly early under conditions of protein malnutrition.

**I. PLASMA PROTEINS MAINTAIN PROPER DISTRIBUTION OF WATER BETWEEN BLOOD AND TISSUES**

When the cells are removed from the blood, the remaining plasma is composed of water, nutrients, metabolites, hormones, electrolytes, and proteins. Plasma has essentially the same electrolyte composition as other extracellular fluids and constitutes approximately 25% of the body’s total extracellular fluid. The plasma proteins serve a number of functions, which include maintaining the proper distribution of water between the blood and the tissues, transporting nutrients, metabolites, and hormones throughout the body, defending against infection, and maintaining the integrity of the circulation through clotting. Many diseases alter the amounts of plasma proteins produced and, hence, their concentration in the blood. These changes can be determined by electrophoresis of plasma proteins over the course of a disease.

**A. Body Fluid Maintenance between Tissues and Blood**

As the arterial blood enters the capillaries, fluid moves from the intravascular space into the interstitial space (that surrounding the capillaries) because of what are known as Starling’s forces. The hydrostatic pressure in the arteriolar end of the capillaries (~37 mm Hg) exceeds the sum of the tissue pressure (~1 mm Hg) and the osmotic pressure of the plasma proteins (~25 mm Hg). Thus, water tends to leave the capillaries and enter extravascular spaces. At the venous end of the capillaries, the hydrostatic pressure falls to approximately 17 mm Hg while the osmotic pressure and the tissue pressure remain constant, resulting in movement of fluid back from the extravascular (interstitial) spaces and into the blood. Thus, most of the force bringing water back from the tissues is the osmotic pressure mediated by the presence of proteins in the plasma.

**B. The Major Serum Protein, Albumin**

As indicated in Table 45.1, the liver produces a number of transport or binding proteins and secretes them into the blood. The major protein synthesized is albumin, which constitutes approximately 60% of the total plasma protein, but because of its relatively small size (69 kDa) is thought to contribute 70 to 80% of the total osmotic pressure of the plasma. Albumin, like most plasma proteins, is a glycoprotein and is a carrier of free fatty acids, calcium, zinc, steroid hormones, copper, and bilirubin.
In spite of the importance of albumin in the maintenance of osmotic pressure in the blood, individuals lacking albumin (analbuminemia) have only moderate edema. Apparently, the concentration of other plasma proteins is increased to compensate for the lack of albumin. The frequency of analbuminemia is less than one per million individuals.

Many drugs also bind to albumin, which may have important pharmacologic implications. For example, when a drug binds to albumin, such binding will likely lower the effective concentration of that drug and may lengthen its lifetime in the circulation. Drug dosimetry may need to be recalculated if a patient’s plasma protein concentration is abnormal.

II. THE PLASMA CONTAINS PROTEINS THAT AID IN IMMUNE DEFENSE

Two different sets of proteins aid in the immune response, the immunoglobulins and the complement proteins. The immunoglobulins are secreted by a subset of differentiated B lymphocytes termed plasma cells and bind antigens at binding sites formed by the hypervariable regions of the proteins (see Chapter 7). Once the antibody–antigen complex is formed, it must be cleared from the circulation. The complement system participates in this function. The complement system consisting of approximately 20 proteins becomes activated in either of two ways. The first is interaction with antigen–antibody complexes, and the second, specific for bacterial infections, is through interaction of bacterial cell polysaccharides with complement protein C3b. Activation of the complement system by either trigger results in a proteolytic activation cascade of the proteins of the complement system, resulting in the release of biologically active peptides or polypeptide fragments. These peptides mediate the inflammatory response, attract phagocytic cells to the area, initiate degranulation of granulocytes, and promote clearance of antigen–antibody complexes.

Protease inhibitors in plasma serve to carefully control the inflammatory response. Activated neutrophils release lysosomal proteases from their granules that can attack elastin, the various types of collagen, and other extracellular matrix proteins. The plasma proteins α1-antiproteinase (α1-antitrypsin) and α2-macroglobulin limit proteolytic damage by forming noncovalent complexes with the proteases, thereby inactivating them. However, the product of neutrophil myeloperoxidase, HOCl, inactivates the protease inhibitors, thereby insuring that the proteases are active at the site of infection.

III. PLASMA PROTEINS MAINTAIN THE INTEGRITY OF THE CIRCULATORY SYSTEM

Blood is lost from the circulation when the endothelial lining of the blood vessels is damaged and the vessel wall is partly or wholly severed. When this occurs, the subendothelial cell layer is exposed, consisting of the basement membrane of the endothelial cells and smooth muscle cells. In response to the damage, a barrier (the hemostatic plug, a fibrin clot), initiated by platelet binding to the damaged area, is formed at the site of injury. Regulatory mechanisms limit clot formation to the site of injury and control its size and stability. As the vessel heals, the clot is degraded (fibrinolysis). Plasma proteins are required for these processes to occur.

Table 45.1. Specific Plasma Binding Proteins Synthesized in the Liver

<table>
<thead>
<tr>
<th>Protein</th>
<th>Function</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ceruloplasmin</td>
<td>Binds copper; appears to be more important as a copper storage pool than as a transport protein; integrates iron and copper homeostasis</td>
</tr>
<tr>
<td>Corticosteroid-binding globulin</td>
<td>Binds cortisol</td>
</tr>
<tr>
<td>Haptoglobin</td>
<td>Binds extracorpuscular heme</td>
</tr>
<tr>
<td>Lipoproteins</td>
<td>Transport cholesterol and fatty acids</td>
</tr>
<tr>
<td>Retinol-binding protein</td>
<td>Binds vitamin A</td>
</tr>
<tr>
<td>Sex hormone–binding globulin</td>
<td>Binds estradiol and testosterone</td>
</tr>
<tr>
<td>Transferrin</td>
<td>Transports iron</td>
</tr>
<tr>
<td>Transthyretin</td>
<td>Binds thyroxine (T4); also forms a complex with retinol-binding protein</td>
</tr>
</tbody>
</table>

α1-Antiproteinase (AAP) is the main serine protease inhibitor of human plasma. Individuals with a point mutation in exon 5 of the AAP gene, which results in a single amino acid substitution in the protein, have diminished secretion of AAP from the liver. These individuals are at increased risk for developing emphysema. When neutrophils degranulate in the lungs as part of the body’s defense against microorganisms, insufficient levels of AAP are present to neutralize the elastase and other proteases released. The excess proteolytic activity damages lung tissue, leading to loss of alveolar elasticity and emphysema. Methionine 358 of AAP is necessary for AAP binding to the proteases. Oxidation of this methionine destroys AAP’s protease-binding capacity. Cigarette smoke oxidizes Met-358, thereby increasing the risk for emphysema.
Von Willebrand factor is a large multimeric glycoprotein with a subunit molecular weight of 220,000 daltons. Its size in the circulation ranges between 500 and 20,000 kDa, and its role in circulation is to stabilize Factor VIII and protect it from degradation. The high-molecular-weight forms are concentrated in the endothelium of blood vessels and are released in response to stress hormones and endothelial damage. High-molecular-weight vWF released by the endothelium is cleaved by a specific metalloprotease in the serum, reducing the size of the circulating vWF. Large vWF multimers are more effective at forming complexes with platelets than are small vWF multimers.

Platelets are derived from megakaryocytes in the bone marrow. Megakaryocytes differentiate from the hemopoietic stem cell. As the megakaryocyte matures, it undergoes synchronous nuclear replication without cellular division, to form a cell with a multilobed nucleus and enlarged cytoplasm. At approximately the 8-nucleus stage, the cytoplasm becomes granular, and the platelets are budded off the cytoplasm. A single megakaryocyte gives rise to approximately 4,000 platelets.

Platelets are non-nucleated cells present in the blood whose major role is to form mechanical plugs at the site of vessel injury and to secrete regulators of the clotting process and vascular repair. In the absence of platelets, leakage of blood from vents in small vessels is common.

In the nonactivated platelet (a platelet not involved in forming a clot), the plasma membrane invaginates extensively into the interior of the cell, forming an open membrane (canalicular) system. Because the plasma membrane contains receptors and phospholipids that accelerate the clotting process, the canalicular structure substantially increases the membrane surface area potentially available for clotting reactions. The platelet interior contains microfilaments and an extensive actin/myosin system. Platelet activation in response to endothelial injury causes Ca\(^{2+}\)-dependent changes in the contractile elements, which, in turn, substantially change the architecture of the plasma membrane. Long pseudopodia are generated, increasing the surface area of the membrane as clot formation is initiated.

Platelets contain three types of granules. The first are electron-dense granules, which contain calcium, adenosine diphosphate (ADP), adenosine triphosphate (ATP), and serotonin. The second type of granule is the α-granule, which contains a heparin antagonist (heparin interferes with blood clotting; see biochemical comments), platelet-derived growth factor, β-thromboglobulin, fibrinogen, von Willebrand factor (vWF), and other clotting factors. The third type of granule is the lysosomal granule, which contains hydrolytic enzymes. During activation, the contents of these granules, which modulate platelet aggregation and clotting, are secreted.

2. PLATELET ACTIVATION

Three fundamental mechanisms are involved in platelet function during blood coagulation: adhesion, aggregation, and secretion. Adhesion sets off a series of reactions termed platelet activation, which leads to platelet aggregation and secretion of platelet granule contents.

The adhesion step refers primarily to the platelet–subendothelial interaction that occurs when platelets initially adhere to the sites of blood vessel injury (Fig. 45.1). Blood vessel injury exposes collagen, subendothelial matrix-bound vWF, and other matrix components. vWF is a protein synthesized in endothelial cells and megakaryocytes, and is located in the subendothelial matrix, in specific platelet granules, and in the circulation bound to Factor VIII. The platelet cell membrane contains glycoproteins (GPs) that bind to collagen and to vWF, causing the platelet to adhere to the damaged vessel wall. This initial adhesion is crucial for platelet activation and subsequent aggregation.

![Fig. 45.1. Adhesion of platelets to the subendothelial cell layer. 1. GPla initially binds to the exposed collagen, which results in changes in the three-dimensional configuration of the complex, allowing GP Ib to bind to vWF (2). 3. This second binding event exposes the GPIIb/GPIIIa complex, which also can bind to vWF and fibrinogen.](image-url)
to adhere to the subendothelium. Binding to collagen by GPIa (integrin α2β1) causes the platelet to change its shape from a flat disc to a spherical cell. The latter extrudes long pseudopods, which promote platelet/platelet interactions. Binding of subendothelial vWF by GPIb causes changes in the platelet membrane that expose GPIIb/IIIa (integrin αIIbβ3) binding sites to fibrinogen and vWF.

The initial adherence of platelets sets off a series of reactions (platelet activation) that results in more platelets being recruited and aggregated at the site of injury. After initial adherence, some of the platelets release the contents of their dense granules and their α granules, with ADP release being of particular importance because ADP is a potent platelet activator. ADP released from the platelets and from damaged red blood cells binds to a receptor on the platelet membrane, which leads to the further unmasking of GPIIb/IIIa binding sites. Aggregation of platelets cannot take place without ADP stimulation, because ADP induces swelling of the activated platelets, promoting platelet/platelet contact and adherence.

Fibrinogen is a protein that circulates in the blood and is also found in platelet granules. It consists of two triple helices held together with disulfide bonds. (Fig.45.2) Binding of fibrinogen to activated platelets is necessary for aggregation,
Thrombotic thrombocytopenic purpura (TTP) is a disease characterized by the formation in the circulation of microclots (microthrombi) consisting of aggregated platelets. The microthrombi collect in the microvasculature and damage red blood cells, resulting in hemolytic anemia. They also damage vascular endothelium, exposing collagen and releasing high-molecular-weight vWF, promoting more platelet aggregation. The subsequent depletion of platelets renders the patient susceptible to internal hemorrhage. Mortality in untreated TTP can approach 90%.

Familial TTP is associated with mutations in the vWF-specific metalloprotease, although not all individuals with defective protease develop TTP. Sporadic cases of TTP are associated with the development of an antibody to the metalloprotease.

In response to collagen and thrombin, platelets release vasoconstrictors. Serotonin is released from the dense granules of the platelets, and the synthesis of thromboxane A2 is stimulated. This will reduce blood flow to the damaged area. Platelet-derived growth factor, which stimulates proliferation of vascular cells, is also released into the environment surrounding the damage.

The utilization of an active site serine to cleave a peptide bond is common to a variety of enzymes referred to as serine proteases. Serine proteases are essential for activating the formation of a blood clot from fibrin. Fibrin and many of the other proteins involved in blood coagulation are present in the blood as inactive precursors orzymogens, which must be activated by proteolytic cleavage. Thrombin, the serine protease that converts fibrinogen to fibrin, has the same aspartic-histidine-serine catalytic triad found in chymotrypsin and trypsin.

Thrombin is activated by proteolytic cleavage of its precursor protein, prothrombin. The sequence of proteolytic cleavages leading to thrombin activation requires Factor VIII, the blood-clotting protein deficient in TTP.

B. The Blood Coagulation Cascade

Thrombin (clot) formation is enhanced by thrombin activation, which is mediated by the complex interaction that constitutes the blood coagulation cascade. This cascade (Fig. 45.3) consists primarily of proteins that serve as enzymes or cofactors, which function to accelerate thrombin formation and localize it at the site of injury. These proteins are listed in Table 45.2. All of these proteins are present in the plasma as proenzymes (zymogens). These precursor proteins are activated by cleavage of the polypeptide chain at one or more sites. The key to successful and appropriate thrombus formation is the regulation of the proteases that activate these zymogens.

The proenzymes (Factors VII, XI, IX, X, XII, and prothrombin) are serine proteases that, when activated by cleavage, cleave the next proenzyme in the cascade. Because of the sequential activation, a great acceleration and amplification of the response is achieved. That cleavage and activation have occurred is indicated by the addition of an “a” to the name of the proenzyme (e.g., Factor IX is cleaved to form the active Factor IXa).

The cofactor proteins (tissue factor, Factors V and VIII) serve as binding sites for other factors. Tissue factor is not related structurally to the other blood coagulation cofactors and is an integral membrane protein that does not require cleavage for active function. Factors V and VIII serve as procofactors, which, when activated by cleavage, function as binding sites for other factors.

Two additional proteins that are considered part of the blood coagulation cascade, protein S and protein C, are regulatory proteins. Only protein C is regulated by proteolytic cleavage, and when activated, is itself a serine protease.

C. The Process of Blood Coagulation

Activation of the blood coagulation cascade is triggered by the reaction of plasma proteins with the subendothelium at the same time that platelets are adhering to the subendothelial layer. Historically, two different pathways were discovered, one dependent on external stimuli (such as blunt trauma, which initiates the extrinsic pathway) and one using internal stimuli (the intrinsic pathway). As our understanding of blood clotting has expanded, it has become obvious that these distinctions are no longer correct, because there is overlap between the pathways, but the terms have persisted in the description of the pathways.

In the case of external trauma, damaged tissues present tissue factor to the blood at the injured site, thereby triggering the extrinsic phase of blood coagulation. Circulating Factor VII binds to tissue factor, which autocatalyzes its own activation to Factor VIIa. Factor VIIa then activates Factor X (to Xa) in the extrinsic pathway and Factor IX (to IXa) in the intrinsic pathway. Factor IXa, as part of the intrinsic pathway, also activates Factor X. Therefore, activation of both the extrinsic and intrinsic pathways result in the conversion of Factor X to Factor Xa. All of these conversions require access to membranes and calcium; the platelet membrane, which had adhered to the damaged site, is used as a scaffold for the activation reactions to occur. The γ-carboxylated clotting proteins are chelated to membrane surfaces via electrostatic interactions with calcium and negatively charged phospholipids of the platelet membrane. The protein cofactors VIIIa and Va serve as sites for assembling enzyme–cofactor complexes on the platelet surface, thereby accelerating and localizing the reaction. The result is thrombin formation, which augments its own formation by converting Factors V, VIII, and XI into activated cofactors and...
Table 45.2. Proteins of Blood Coagulation

<table>
<thead>
<tr>
<th>Factor</th>
<th>Descriptive Name</th>
<th>Function/Active Form</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>Fibrinogen</td>
<td>Fibrin</td>
</tr>
<tr>
<td>II</td>
<td>Prothrombin</td>
<td>Serine protease</td>
</tr>
<tr>
<td>III</td>
<td>Tissue factor</td>
<td>Receptor and cofactor</td>
</tr>
<tr>
<td>IV</td>
<td>(\text{Ca}^{2+})</td>
<td>Cofactor</td>
</tr>
<tr>
<td>V</td>
<td>Proaccelerin, labile factor</td>
<td>Cofactor</td>
</tr>
<tr>
<td>VII</td>
<td>Proconvertin</td>
<td>Serine protease</td>
</tr>
<tr>
<td>VIII</td>
<td>Antihemophilia factor A</td>
<td>Cofactor</td>
</tr>
<tr>
<td>IX</td>
<td>Antihemophilia factor B, Christmas factor</td>
<td>Serine protease</td>
</tr>
<tr>
<td>X</td>
<td>Stuart-Prower factor</td>
<td>Serine protease</td>
</tr>
<tr>
<td>XI</td>
<td>Plasma thromboplastin antecedent</td>
<td>Serine protease</td>
</tr>
<tr>
<td>XII</td>
<td>Hageman (contact) factor</td>
<td>Serine protease</td>
</tr>
<tr>
<td>XIII</td>
<td>Fibrin stabilizing factor</td>
<td>(\text{Ca}^{2+})-dependent transglutaminase</td>
</tr>
<tr>
<td>Prekallikrein</td>
<td></td>
<td>Serine protease</td>
</tr>
<tr>
<td>High-molecular-weight kininogen</td>
<td></td>
<td>Cofactor</td>
</tr>
</tbody>
</table>

**Table 45.2. Proteins of Blood Coagulation**

**Coagulation Factors**

**Regulatory Proteins**

<table>
<thead>
<tr>
<th>Factor</th>
<th>Function/Active Form</th>
</tr>
</thead>
<tbody>
<tr>
<td>Thrombomodulin</td>
<td>Endothelial cell receptor, binds thrombin</td>
</tr>
<tr>
<td>Protein C</td>
<td>Activated by thrombomodulin-bound thrombin; is a serine protease</td>
</tr>
<tr>
<td>Protein S</td>
<td>cofactor; binds activated protein C</td>
</tr>
</tbody>
</table>

Fig. 45.3. Blood coagulation cascade. Activation of clot formation occurs through two separate but interlocking pathways, termed the intrinsic and extrinsic pathways. The intrinsic pathway is activated when plasma proteins react with the exposed subendothelium of the damaged blood vessel. Platelets and vWF bind to the exposed subendothelium, and the platelets, in turn, bind fibrinogen. The extrinsic pathway is activated by tissue factor. The reactions designated by “PL, Ca” are occurring through cofactors bound to phospholipids (PL) on the cell surface in a \(\text{Ca}^{2+}\)-coordination complex. Factors XIIa, XIa, IXa, VIIa, Xa, and thrombin are serine proteases. Note the positive feedback regulation of thrombin on the activation of proteases earlier in the cascade sequence. HMWK = high-molecular-weight kininogen.
The initial activation of prothrombin to thrombin is slow, because the activator cofactors, Factors VIIIa and Va, are only present in small amounts. However, once a small amount of thrombin is activated, it will accelerate its own production by cleaving Factors V and VIII to their active forms.

stimulating platelet degranulation. Note that these factors are in the intrinsic pathway. The intrinsic pathway is thought to sustain the coagulation response initiated by the extrinsic pathway. The major substrate of thrombin is fibrinogen, which is hydrolyzed to form fibrin monomers that undergo spontaneous polymerization to form the fibrin clot. This is considered a “soft” clot because the fibrin monomers are not cross-linked. Cross-linking requires Factor XIIIa, which is activated by thrombin cleavage of Factor XIII.

1. CROSS-LINKING OF FIBRIN

Factor XIIIa catalyzes a transamidation reaction between Gln and Lys side chains on adjacent fibrin monomers. The covalent cross-linking takes place in three dimensions, creating a strong network of fibers resistant to mechanical and proteolytic damage. This network of fibrin fibers traps the aggregated platelets and other cells, forming the clot that plugs the vent in the vascular wall. (Fig. 45.4) Factor XIIIa is the only enzyme in the blood coagulation cascade that is not a serine protease.

2. KALLIKREIN AND HIGH-MOLECULAR-WEIGHT KININOGEN (HMWK)

The classical intrinsic pathway begins with the assembly of prekallikrein, high-molecular-weight kininogen (HMWK), Factor XII, and Factor XI on a negatively charged surface, presumably an endothelial cell in vivo (see Fig. 45.3). High-molecular-weight kininogen is a glycoprotein that binds prekallikrein and aids in its assembly on the endothelial cell. Prekallikrein is the zymogen form of a serine protease. Factor XII autoactivates, forming Factor XIIa, which converts prekallikrein to kallikrein. Kallikrein then enhances the activation of Factor XII, which leads to the activation of Factors XI and VII.

How important these steps are in the initiation of the coagulation cascade is unknown. Individuals lacking HMWK, prekallikrein, or Factor XII do not suffer from bleeding disorders. Under usual conditions, activation of Factor VII with subsequent activation of Factors IX and X is thought to be sufficient to activate the coagulation pathway.

3. FACTOR COMPLEXES

In several of the essential steps in the blood coagulation cascade, the activated protease is bound in a complex attached to the surface of the platelets that have aggregated at the site of injury. Factors VII, IX, X, and prothrombin contain a domain in which one or more glutamate residues are carboxylated to γ-carboxyglutamate in a reaction requiring vitamin K (Fig 45.5). Prothrombin and Factor X both contain 10 or more γ-carboxyglutamate residues that bind Ca^{2+}. Ca^{2+} forms a coordination complex with the negatively charged platelet membrane phospholipids and the γ-carboxyglutamates, thereby localizing the complex assembly and thrombin formation to the platelet surface.

Cofactor Va contains a binding site for both Factor Xa and prothrombin, the zymogen substrate of Factor Xa. On binding to the Factor Va–platelet complex, prothrombin undergoes a conformational change, rendering it more susceptible to enzymatic cleavage. Binding of Factor Xa to the Factor Va–prothrombin–platelet complex allows the prothrombin-to-thrombin conversion. Complex assembly accelerates the rate of this conversion 10,000- to 15,000-fold as compared with non–complex formation.

Factor VIIIa forms a similar type of complex on the surface of activated platelets, but binds Factor IXa and its zymogen substrate, Factor X. Tissue factor works slightly differently because it is an integral membrane protein. However, once exposed by injury, it also binds Factor VIIa and initiates complex formation.
Complex assembly has two physiologically important consequences. First, it enhances the rate of thrombin formation by as much as several hundred thousand-fold, enabling the clot to form rapidly enough to preserve hemostasis. Secondly, such explosive thrombin formation is localized to the site of vascular injury at which the negatively charged phospholipids are exposed. From our knowledge of the location of such phospholipids in cellular and subcellular organelle membranes, these surface-binding sites are only exposed at an injury site in which cell rupture exposes the internal membrane surface (recall that certain phospholipids only face the cytoplasm; if these lipids are now exposed to the environment, substantial cell damage must have occurred).

4. VITAMIN K REQUIREMENT FOR BLOOD COAGULATION

The formation of the \(\gamma\)-carboxyglutamate residues on blood coagulation factors takes place in the hepatocyte before release of the protein. Within the hepatocyte, vitamin K (which is present in the quinone form) is reduced to form vitamin KH\(_2\) by a microsomal quinone reductase (see Fig.45.5). Vitamin KH\(_2\) is a cofactor for carboxylases that add a carboxyl group to the appropriate glutamate residues in the proenzyme to form the carboxylated proenzyme (e.g., prothrombin). In the same

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**Fig. 45.5.** A. Structures of vitamin K derivatives. Phylloquinone is found in green leaves, and intestinal bacteria synthesize menaquinone. Humans will convert menadione to a vitamin K active form. B. Vitamin K–dependent formation of \(\gamma\)-carboxyglutamate residues. Thrombin, Factor VII, Factor IX, and Factor X are bound to their phospholipid activation sites on cell membranes by Ca\(^{2+}\). The vitamin K–dependent carboxylase, which adds the extra carboxyl group, uses a reduced form of vitamin K (KH\(_2\)) as the electron donor and converts vitamin K to an epoxide. Vitamin K epoxide is reduced, in two steps, back to its active form by the enzymes vitamin K epoxide reductase and vitamin K reductase.
Warfarin (Coumadin®) is a slow-and long-acting blood anticoagulant with a structure resembling that of vitamin K. The structural similarity allows the compound to compete with vitamin K and prevent γ-carboxylation of glutamate residues in Factors II, VII, IX, X, and proteins C and S. The noncarboxylated blood clotting protein precursors increase in both the blood and plasma, but they are unable to promote blood coagulation because they cannot bind calcium and thus cannot bind to their phospholipid membrane sites of activation.

Deficiency in the amount or functionality of protein C or protein S increases the risk for venous thromboembolism. Homozygous individuals for these mutations do not survive the neonatal period unless given replacement therapy.

In European populations, a point mutation in the Factor V gene (Factor V Leiden) causes the replacement of an arg with a gln in the preferred site for cleavage by activated protein C, rendering Factor Va Leiden resistant to APC. Heterozygous individuals have a sixfold to eightfold increased risk of deep vein thromboses, and homozygous individuals have a 30- to 140-fold increased risk. The Factor V Leiden mutation does not appear to be associated with increased risk for arterial thrombosis, such as myocardial infarction, except in young women who smoke.

Genetic studies suggest that the Factor V Leiden mutation arose after the separation of the European, Asian, and African populations. The frequency of this variant indicates that it conferred some selective advantage at one time. In the developed world, inherited APC resistance is the most prevalent risk factor for familial thrombotic disease.

Fig. 45.6. Anti-thrombotic effects of thrombin. Thrombin, bound to thrombomodulin on the endothelial cell surface, activates protein C. Activated protein C, in complex with protein S, binds to the platelet membrane, and the activated complex destroys Factors Va and VIIIa, thereby inhibiting the coagulation cascade.
mechanisms control and limit intravascular proteolysis. The serpins (serine protease inhibitors) are a group of naturally occurring inhibitory proteins present in the plasma at high concentration (approximately 10% of the plasma proteins are serpins). Eight major inhibitors have been found that share a common mechanism of action and inhibit proteases involved in coagulation and clot dissolution (fibrinolysis). Each inhibitor possesses a reactive site that appears to be an ideal substrate for a specific serine protease and thus acts as a trap for that protease. The bound serine protease attacks a peptide bond located at a critical amino acid residue within the serpin and forms a tight enzyme–inhibitor complex.

The activity of thrombin is controlled by the serpin antithrombin III (ATIII). Regulation of blood coagulation at the level of thrombin is critical because this enzyme affects the pathways of both coagulation and fibrinolysis (see section F). One molecule of ATIII irreversibly inactivates one molecule of thrombin through reaction of an arginine residue in ATIII with the active-site serine residue of thrombin. ATIII–thrombin complex formation is markedly enhanced in the presence of heparin. Heparin is a glycosaminoglycan (see Chapter 49) found in the secretory granules of mast cells and in the loose connective tissue around small vascular beds. Heparin binds to lysyl residues on ATIII and dramatically accelerates its rate of binding to thrombin. This is because of an allosteric alteration in ATIII such that the position of the critical arginine residue of ATIII is more readily available for interaction with thrombin. The formation of the ATIII–thrombin complex releases the heparin molecule so that it can be reused, and therefore, the function of heparin is catalytic. Thrombin that is attached to a surface, for example, to thrombomodulin on the endothelial cell membrane, is no longer participating in clot formation and is not readily attacked by ATIII or the ATIII–heparin complex. The ATIII–heparin complex also can inactivate the serine protease Factors XIIIa, XIa, IXa and Xa, but has no effect on Factor VIIa or activated protein C.

E. Thromboresistance of Vascular Endothelium

Endothelial cells of blood vessels provide a selectively permeable barrier between the circulating blood and the tissues. The normal endothelial cell lining neither activates coagulation nor supports platelet adhesion; thus, it is called a nonthrombogenic surface. The thromboresistance of normal vascular endothelium is contributed to by several properties. Endothelial cells are highly negatively charged, a feature that may repel the negatively charged platelets. Endothelial cells synthesize prostaglandin I2 (PGL2) and nitric oxide, vasodilators and powerful inhibitors of platelet aggregation. PGI2 synthesis is stimulated by thrombin, epinephrine, and local vascular injury. Endothelial cells also synthesize two cofactors that each inhibit the action of thrombin, thrombomodulin and heparan sulfate. Heparan sulfate is a glycosaminoglycan similar to heparin that potentiates antithrombin III, but not as efficiently. The inactivation of thrombin is accelerated by heparan sulfate present on the endothelial cell surface. Thus, the intact endothelium has the capability of modifying thrombin action and inhibiting platelet aggregation.

F. Fibrinolysis

After successful formation of a hemostatic plug, further propagation of the clot must be prevented. This is accomplished in part by switching off blood coagulation and in part by turning on fibrinolysis. Fibrinolysis involves the degradation of fibrin in a clot by plasmin, a serine protease that is formed from its zymogen, plasminogen. Plasminogen is a circulating serum protein that has a high affinity for fibrin, promoting the incorporation of plasminogen in the developing clot. The activity of plasminogen is mediated by proteins known as plasminogen activators. The conversion of plasminogen to plasmin by plasminogen activators can occur both in the liquid phase of the blood and at the clot surface; however, the latter process is by
Fig. 45.7. Regulation of plasmin activation. Plasminogen can be activated by either t-PA or scu-PA (+). PAI-1 blocks t-PA action (−). Streptokinase binding to plasminogen allows autocatalysis to form plasmin. Circulating α2-antiplasmin blocks (−) the activity of any soluble plasmin that may be in the blood.

Both streptokinase and t-PA have been approved for the treatment of myocardial infarction. Both reduce mortality. Although there are more side effects associated with the use of streptokinase, it is substantially cheaper than t-PA.

far more efficient. Activated protein C (APC), in addition to turning off the blood coagulation cascade, also stimulates the release of plasminogen activator from tissues (t-PA, tissue plasminogen activator) and simultaneously inactivates an inhibitor of plasminogen activator, PAI-1.

Plasminogen activator release can lead to plasmin formation in the circulation. However, the circulating plasmin is rapidly inactivated by binding with α2-antiplasmin, a circulating protease inhibitor. Clot-bound plasmin is not readily inactivated by α-antiplasmin. Thus, plasminogen binding to fibrin facilitates its activation to plasmin, protects it from blood serpins, and localizes it on the fibrin substrate for subsequent efficient proteolytic attack. This mechanism allows for dissolution of fibrin in pathologic thrombi or oversized hemostatic plugs, and at the same time prevents degradation of fibrinogen in the circulating blood.

Two endogenous plasminogen activators are most important; both are synthesized in a variety of cells. Tissue plasminogen activator (t-PA) is chiefly produced by the vascular endothelial cells, has a high binding affinity for fibrin, and plays a major role in fibrinolysis. Single-chain urokinase (scu-PA), is synthesized in most cells and tissues and has a moderate affinity for fibrin. Streptokinase, the bacterial exogenous plasminogen activator from β-hemolytic streptococci, is not an enzyme but an allosteric modifier of human plasminogen that allows an autocatalytic reaction such that plasmin is formed. In vivo, physical stress, hypoxia, and large numbers of low-molecular-weight organic compounds promote increased synthesis and release of t-PA and scu-PA from tissues into the blood. The balance between release of plasminogen activators, the availability of fibrin, and inhibitors of the activators and plasmin determines regulation of the fibrinolytic response, as indicated in Figure 45.7.

G. Regulation of Fibrinolysis

Anti-activators regulate interaction of plasminogen in blood with plasminogen activators in a dynamic equilibrium. Even if minute amounts of plasmin are generated (e.g., after release of vascular plasminogen activator after stress), the enzyme is probably inactivated by antiplasmin. On activation of the blood coagulation system, a fibrin clot is formed, which not only strongly binds t-PA and plasminogen from blood but also accelerates the rate of plasmin activation. The clot-bound plasmin is protected from inhibitors while attached to fibrin. The enzyme is inactivated by α2-antiplasmin and α2-macroglobulin after proteolytic dissolution of fibrin and its liberation into the liquid phase of blood. Thus, the fibrin network catalyzes both initiation and regulation of fibrinolysis.

CLINICAL COMMENTS

Sloe Klotter has hemophilia A, the most frequently encountered serious disorder of blood coagulation in humans, occurring in 1 in every 10,000 males. The disease is transmitted with an X-linked pattern of inheritance.

The most common manifestations of hemophilia A are those caused by bleeding into soft tissues (hematomas) such as muscle or into body spaces such as the peritoneal cavity or the lumen of the gastrointestinal tract. When bleeding occurs repeatedly into joints (hemarthrosis), the joint may eventually become deformed and immobile.

In the past, bleeding episodes have been managed primarily by administration of Factor VIII, sometimes referred to as antihemophilia cofactor. Unfortunately, the concentration of Factor VIII in plasma is quite low (0.3 nM compared with 8,800 nM for fibrinogen), requiring that it be prepared from multiple human donors. Before donor screening and virus inactivation procedures during preparation...
Another X-linked bleeding disorder is hemophilia B, which is caused by mutations in the gene for Factor IX. Lack of Factor IX activity leads to an inability to convert prothrombin to thrombin, and impaired clotting.

BIOCHEMICAL COMMENTS

A number of drugs have been developed that inhibit the blood coagulation cascade. Such drugs are useful in cases in which patients develop spontaneous thrombi, which, if left untreated, would result in a fatal pulmonary embolism. There are three major classes of such drugs; the heparins, vitamin K antagonists, and specific inhibitors of thrombin.

Heparin will bind to and activate ATIII, which leads to thrombin inactivation. ATIII also blocks the activity of Factors VIIIa, IXa, Xa, and XIa. Heparin can be administered in either of two forms: unfractionated, or high-molecular-weight (HMW) heparin, and fractionated, or low-molecular-weight (LMW) heparin. HMW heparin is a heterogeneous mixture of glycosaminoglycans, with an average chain length of 45 monosaccharides with an average molecular weight of 15 kDa (the range is 3–30 kDa). LMW heparins are fragments of HMW heparin, containing fewer than 18 monosaccharides with an average molecular weight of 4 to 5 kDa.

HMW heparin will bind to plasma proteins and cell surfaces in addition to its prime target, ATIII. Because different individuals synthesize different levels of plasma proteins, the use of this form of heparin as an anticoagulant requires constant monitoring of the patient to ensure that the correct dosage has been given such that spontaneous thrombi do not develop, but not so much that spontaneous bleeding occurs. LMW heparin has fewer nonspecific interactions than HMW heparin, and its effects are easier to predict on patients, so that constant monitoring is not required.

A major complication of heparin therapy is heparin-induced thrombocytopenia (HIT, excessive blood clotting with a reduction in the number of circulating platelets). This unexpected result of heparin treatment is caused by heparin binding to a platelet protein, platelet factor 4 (PF4), which induces a conformational change in PF4 such that the immune system believes the complex is foreign. Thus, antibodies are developed against the heparin–PF4 complex. When the antibodies bind to the platelets, the platelets become activated, and thrombi develop. Treatment consists of removing the heparin and using a different form of anti-thrombotic agent.

The classic vitamin K antagonist is warfarin. Warfarin acts by blocking the vitamin K reductase enzymes required to regenerate active vitamin K (see Fig. 45.5). This results in reduced γ-carboxylation of Factors II, VII, IX, and X. In the absence of γ-carboxylation, the factors cannot bind calcium nor form the complexes necessary for the coagulation cascade to be initiated. However, warfarin also blocks the activity of proteins S and C, so both blood clotting and the regulation of clotting are impaired by warfarin administration.

Both heparin and warfarin therapy suffer from their lack of specificity, so drugs specific for single steps in the blood coagulation pathway have been sought and identified. Analysis of heparin potentiation of Factor Xa binding to ATIII showed that a unique pentasaccharide sequence was required. An appropriate pentasaccharide, named fondaparinux, was developed that would specifically enhance ATIII interactions with Factor Xa (Fig. 45.8). Fondaparinux stimulates the binding of ATIII to Factor Xa by 300-fold and is specific for Factor Xa inhibition. Fondaparinux does not affect thrombin or platelet activity, and it is not an activating agent of platelets. Because fondaparinux does not bind to PF4, HIT is not a complication with this therapy.
The edema observed in patients with non-calorie protein malnutrition is due to which of the following?

(A) Loss of muscle mass
(B) Ingestion of excess carbohydrates
(C) Increased fluid uptake
(D) Reduced protein synthesis in the liver
(E) Increased ketone body production

Direct thrombin inhibitors are based on the hirudin molecules, which were initially discovered in leeches and other blood-sucking organisms. These organisms would not be able to feed if the blood clotted at the site of the puncture wound, so the organisms secrete thrombin inhibitors to prevent clotting from occurring. Hirudin treatment itself is dangerous in that formation of the hirudin–thrombin complex is irreversible, and use of the drug requires constant monitoring of the patient. Thus, to overcome this problem, rational drug design based on the hirudin structure was used, and a synthetic 20–amino acid peptide known as bivalirudin was synthesized. This agent has a high binding affinity and specificity for thrombin although its effects on thrombin are transient (not irreversible), making this a safer agent for long-term use.

Suggested References


REVIEW QUESTIONS—CHAPTER 45

1. The edema observed in patients with non-calorie protein malnutrition is due to which of the following?

   (A) Loss of muscle mass
   (B) Ingestion of excess carbohydrates
   (C) Increased fluid uptake
   (D) Reduced protein synthesis in the liver
   (E) Increased ketone body production
2. A recent surgery patient receiving warfarin therapy was found to be bleeding internally. The clotting process is impaired in this patient primarily because of which of the following?

(A) Inability of the liver to synthesize clotting factors
(B) A specific inhibition of Factor XIII activation
(C) An inability to form clotting factor complexes on membranes
(D) A reduction of plasma calcium levels
(E) An enhancement of protein C activity

3. An inactivating mutation in which of the following proenzymes would be expected to lead to thrombosis, uncontrolled blood clotting?

(A) Factor XIII
(B) Prothrombin
(C) Protein C
(D) Factor VIII
(E) Tissue factor

4. Classical hemophilia A results in an inability to directly activate which of the following factors?

(A) Factor II
(B) Factor IX
(C) Factor X
(D) Protein S
(E) Protein C

5. Hemophilia B results in an inability to directly activate which of the following factors?

(A) Factor II
(B) Factor IX
(C) Factor X
(D) Protein S
(E) Protein C